

AviRex™ Plant RNA

for plant tissue and cell culture

Before Starting

Add 48 ml of absolute ethanol to the PW (only at the first use).

Reagents NOT Provided

- Chloroform
- 2. 70% and 96% ethanol

Protocol

- Cutting the tissue into the small pieces on a sterile petri dish by a scalpel and grind with a mortar and pestle under liquid nitrogen. Transfer 50-100 mg of tissue or 6 x 10° cells (for cell cultures) into a 1.5 ml tube and add 800 µl of RL solution.
- Pipetting the tissue into and out of the tip to avoid clumps. You can also homogenize hard tissue by homogenizer on ice. Incubate at room temperature for 8 min.
- Add 200 µl of chloroform to the mixture. Shake it completely for 15 s and incubate for 3 min at room temperature.
- Spin for 12 min at 13,000 rpm at 4 °C.
- Transfer 400 μl of the upper phase into a new 1.5 ml tube. Add equal Volume of 70% ethanol (use 96% ethanol for low RNA samples) to the mixture and mix them well.
 - Transfer mixture to the spin column. DO NOT touch upper rim of column. spin for 1 min at 13.000 rpm.
 - Pour off the flow-through of collection tube.
 - 8 Add 700 μl of PW and spin for 1 min at 13.000 rpm.
 - 9 Pour off the flow-through of collection tube. (Optional: repeat step 8 and 9 with 500 μl of PW to have more pure RNA)
- Spin for 2 min at 13.000 rpm to remove the remaining of the wash buffer . Transfer the spin column to a new 1.5 ml microtube.
 - Add 50 µl of DEPC-treated water, wait 3 min at room temperature. if you want more concentration add less DEPC-treated water (35 µl).
 - Spin for 1 min at 13.000 rpm to elute RNA from the column. Store RNA solution at -70 C.







400 µl of upper phase Add 400 µl ethanol



700 µl PW Spin 1 min





Spin 1 mir



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kit content			
No.	Description:	Packing	Qty.
1	RL Buffer (RNA Lysis Buffer)	20 ml	2
2	PW Buffer (Wash Buffer)	12 ml	1
3	DEPC-treated Water	3 ml	1
4	Spin Column	50	1
5	Collection Tube	50	1

Research Use Only

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